

# Investigation of Intestine Bacterial Abundance Change and Mortality Rate of Waxworms (Galleria mellonella. L)

after Consuming Polystyrene



Author: Ruo-shi, Wang Instructor: Jun, Chen

School: TAIPEI MUNICIPAL LISHAN HIGH SCHOOL

#### Introduction

#### I. Motivation

Plastic pollution is a severe issue, and biodegradation is considered a potential solution. It is known that certain microorganisms, such as the intestine bacteria of waxworms (Galleria mellonella. L) can degrade polystyrene (PS).

#### **Known facts**

- Waxworms have the ability to degrade PS.
- Waxworms' mortality rate increases after they degrade PS.
- Waxworms exhibit an increase in intestinal bacterial abundance after consuming PS.

#### II. Purpose

- A. To investigate the mortality rate of the waxworms after their plastic consumption under different temperature settings.
- B. To discuss the correlation between the changes in intestine bacterial number of the waxworms and the increased mortality rate after plastic consumption.
- C. To understand the growth cycle of the waxworms in different temperatures.



Fig 1: Waxworms (Galleria mellonella, L)

#### **III. Experimental Species**

Waxworm is the larval stage of the greater wax moth. Since the intestine bacteria species of the waxworms increase rather than decrease after consuming polystyrene (PS) (Jiang S, et al, 2021), it was chosen as the experimental subject.

The optimal temperature range for waxworm to grow is between 29°C and 35°C, and temperatures between 10°C and 16°C can affect its development. Therefore, 30°C and 15°C were determined to be the temperatures for two different experiment settings.

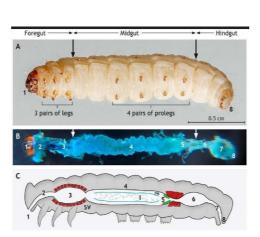


Fig 2: The intestine of the waxworm (Philipp-Albert Sänger, et al. (2022))

## **Procedure**



Weight

Remove Waxworms' feces

Remove dead waxworm

every other day

everyday

everyday





On the 15th day

Weight the feed Take out the left intestine from the waxworms

Measure the absorbance of waxworm intestine bacteria sample

Fig 6: 15°C Experiment process

Group 30°C

Went through same experiment procedures except taking out the intestine from the waxworms on the 10<sup>th</sup> day, due to the faster growth rate of waxworms.

#### **Results and Discussion**

### I. Average mortality rate in different temperatures

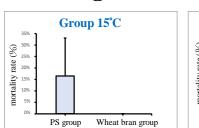
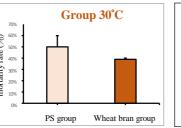


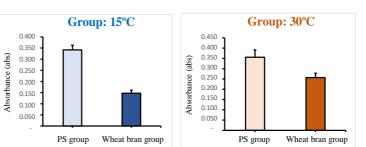
Fig 7: Waxworm's average mortality rate (%)



The average mortality rate of waxworm(%):

- 1. PS group > Wheat bran group
- 2.  $30 \,^{\circ}\text{C} \, \text{group} > 15 \,^{\circ}\text{C} \, \text{group}$

## II. The average absorbance values of intestine bacteria in different temperatures



The average bacterial count in the intestine of the waxworm:

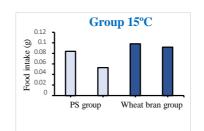
PS group > Wheat bran group

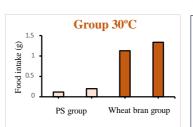
0.2570 .570x108

Fig 8: The average absorbance values of intestine bacterial at different temperatures

5		'	Bacterial: 10 <sup>8</sup> amou		
Group: 15°C					Group: 30°C
Groups	PS group	Wheat bran group	Groups	PS group	Wheat bra
Absorbance (abs)	0.3420	0.1475	Absorbance (abs)	0.3555	
Bacterial count (amount/ml)	$3.420 \times 10^8$	$1.475 \times 10^{8}$	Bacterial count (amount/ml)	$3.555 \times 10^{8}$	2.5

#### III. The food intake of the waxworm in different temperatures





The food intake of waxworms:

- 1. Wheat bran group > PS group
- 2. 30 °C group > 15 °C group

Fig 9:Food intake (g)

#### IV. Average weight changes of waxworm on the 10th day

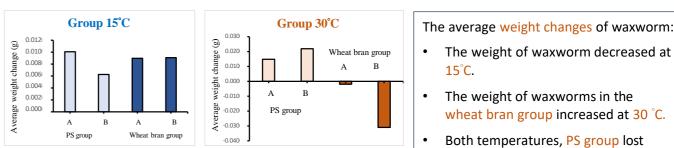


Fig 10: Average weight changes between 0 to 10th day(0d-10d) (g)

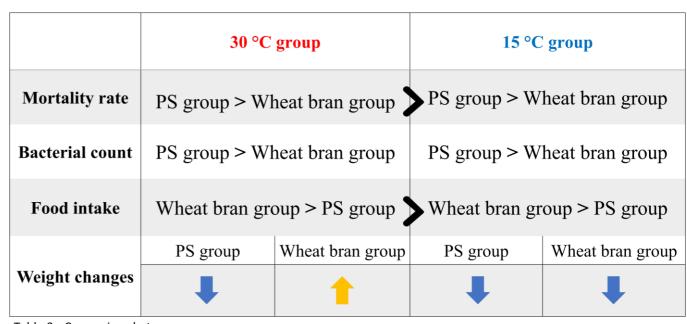
- The weight of waxworm decreased at
- The weight of waxworms in the wheat bran group increased at 30 °C.
- Both temperatures, PS group lost

	30 °C group	15 °C group
Polystyrene	weight decreased	weight decreased
Wheat Bran Main food source	weight increased	weight decreased

Table 1: Weight changes in different group

**Conclusions** 

This experiment shows a new evidence that the total intestine bacterial count increased when the waxworm consumed PS, suggesting that it might be a contributing factor to the mortality rate.



#### Table 2: Comparison between groups

**Suggestions** 

In addition to measuring the bacterial count with spectrophotometer, I also want to use gas chromatography to measure the PS content in waxworm. Moreover, I hope to further investigate the diversity of intestine bacteria by DNA sequencing.

#### References

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